

## RESEARCH OUTPUTS / RÉSULTATS DE RECHERCHE

### Community- and ecosystem-level effects of multiple environmental change drivers

DE LAENDER, Frederik

*Published in:*  
Global Change Biology

*DOI:*  
[10.1111/gcb.14382](https://doi.org/10.1111/gcb.14382)

*Publication date:*  
2018

*Document Version*  
Peer reviewed version

[Link to publication](#)

*Citation for published version (HARVARD):*

DE LAENDER, F 2018, 'Community- and ecosystem-level effects of multiple environmental change drivers: Beyond null model testing', *Global Change Biology*, vol. 24, no. 11, pp. 5021-5030.  
<https://doi.org/10.1111/gcb.14382>

#### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

#### Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

**Title: Community- and ecosystem-level effects of multiple environmental change drivers:  
beyond null model testing**

**Running head: From null models to theory-based predictions**

**Author and affiliation:** Frederik De Laender, Research Unit in Environmental and  
Evolutionary Biology, Namur Institute of Complex Systems, and the Institute of Life, Earth,  
and Environment, University of Namur, Belgium. T: +32478429761;  
[Frederik.delaender@unamur.be](mailto:Frederik.delaender@unamur.be)

**Keywords:** Community ecology, ecosystems, theoretical ecology, multiple stressors,  
environmental stress, eutrophication, resource ratio theory, traits

**Paper type:** Opinion

## **Abstract:**

Understanding the joint effect of multiple drivers of environmental change is a key scientific challenge. The dominant approach today is to compare observed joint effects with predictions from various types of null models. Drivers are said to combine synergistically (antagonistically) when their observed joint effect is larger (smaller) than that predicted by the null model. Here, I argue that this approach does not promote understanding of effects on important community- and ecosystem-level variables such as biodiversity and ecosystem function. I use ecological theory to show that different mechanisms can lead to the same deviation from a null model's prediction. Inversely, I show that the same mechanism can lead to different deviations from a null model's prediction. These examples illustrate that it is not possible to make strong mechanistic inferences from null models. Next, I present an alternative framework to study such effects. This framework makes a clear distinction between two different kinds of drivers (resource ratio shifts and multiple stressors) and integrates both by incorporating stressor effects into resource uptake theory. I show that this framework can advance understanding because of three reasons. First, it forces formalisation of "multiple stressors", using factors that describe the number and kind of stressors, their selectivity and dynamic behaviour, and the initial trait diversity and tolerance among species. Second, it produces testable predictions on how these factors affect biodiversity and ecosystem function, alone and in combination with resource ratio shifts. Third, it can fail in informative ways. That is, its assumptions are clear, so that different kinds of deviations between predictions and observed effects can guide new experiments and theory improvement. I conclude that this framework will more effectively progress understanding of global change effects on communities and ecosystems than does the current practice of null model testing.

## **Null models to study the joint effects of multiple environmental change drivers**

Ecosystems today are invariably challenged by a multitude of environmental change drivers (Brook *et al.*, 2008; Crain *et al.*, 2008; Halpern *et al.*, 2015; Halpern *et al.*, 2008; Schäfer *et al.*, 2016). Understanding their joint effect is a pressing scientific and societal need (Côté *et al.*, 2016; Dupont & Pörtner, 2013; Rudd, 2014). During past years, substantial effort has been put in quantifying such joint effects, with **many** studies focusing at the physiological level (Brennan & Collins, 2015; Gunderson *et al.*, 2016; Häder & Gao, 2015; Sokolova, 2013), while **fewer** have addressed how the effects of individual drivers scale up to affect higher levels of biological organisation such as communities and ecosystems (Garnier *et al.*, 2017).

The dominant approach to examine effects of multiple drivers is to use null models. These null models predict joint effects based on the effects elicited by the individual drivers. When the observed joint effects are smaller than or greater than those predictions, so-called ‘antagonistic’ or ‘synergistic’ effects are concluded, respectively (Crain *et al.*, 2008). A variety of null models exists, each with their own assumptions and limitations (Piggott *et al.*, 2015), and comprehensive overviews exist in the literature (Schäfer & Piggott, 2018). In general, these null models differ in their underlying assumption on how drivers combine to affect the biological variable of interest. The most frequently used null model is additive: it assumes that the effects of the individual drivers add up, correcting for non-independence among drivers if needed. The null model approach is applied to study effects at all levels of biological organisation, going from the individual to the community-level, and are used in both primary studies and meta-analyses (Crain *et al.*, 2008; Darling & Cote, 2008; Jackson *et al.*, 2016; Tolkkinen *et al.*, 2015).

**Null models do not advance comprehension of community and ecosystem-level effects.**

The application of null model testing to community- and ecosystem-level effects of multiple drivers has been criticized before (Kroeker *et al.*, 2017; Thompson *et al.*, 2018). Most of these critiques revolve around the fact that drivers combine differently at different levels of biological organisation (Schäfer & Piggott, 2018). For example, additivity of effects at the level of populations does not automatically imply additivity at the community level (Kroeker *et al.*, 2017). To remediate this difficulty, the development of new null models has been recently proposed (Thompson *et al.*, 2018). In this opinion, however, I propose an alternative solution: to move away from testing null models on community and ecosystem data. My main argument to do so is that current null model testing at the level of communities and ecosystems does not advance comprehension. While null model testing has certainly facilitated meta-analyses, it has until now not augmented general insight in the mechanisms linking environmental change, multiple drivers, and biodiversity and ecosystem function (De Laender *et al.*, 2016; Griffen *et al.*, 2016; Jackson *et al.*, 2016).

The premise of my argument is that null models can only advance comprehension when deviations from their predictions advance mechanistic insight. That is, deviations from their predictions should point towards implications other than “the null model is incorrect”. An example of a null model advancing comprehension, taken from biodiversity science, is the null model of biodiversity effects on ecosystem function (Baert *et al.*, 2017; Fox, 2005; Loreau & Hector, 2001). This null model is based on a clear and explicit assumption: interactions within species (i.e. among conspecifics) are equal to interactions among species (i.e. among heterospecifics). Given this assumption, it is mathematically inevitable that ecosystem functions stay constant with the number of species in the community. Exactly because of the clarity of the underlying assumption, deviations from the null model’s predictions are

informative: they point towards interactions within and between species that are of unequal strength.

Null models in multiple stressor and environmental change research make clear predictions (e.g. effects add up when the model is additive). However, at the community- and ecosystem-level, it is not clear on which hypothesis these predictions are based (Schäfer & Piggott, 2018). Rather, the hypothesis seems to equal the prediction, which hampers inference and thus scientific progress. Indeed, deviations from null model predictions only imply that the joint effect is, for example, not additive. In what follows, I illustrate this point using classic theory in community ecology and the case of species richness as an example of a community-level variable. I do so by showing that there is no link between the sign and size of the deviations from a null model and the community-level mechanisms causing these deviations. More specifically, I show that the same mechanism can lead to different deviations (antagonism and synergism), but also that the opposite holds true: two inherently different mechanisms can both lead to the same deviation (e.g. antagonism).

I consider two species X and Y competing for two essential nutrients  $R_1$  and  $R_2$ , according to resource uptake theory (Chase & Leibold, 2003; Tilman, 1982), assuming Monod uptake and optimal foraging (Tilman, 1982). In this model formulation, a species' competitive ability for  $R_j$  is greater when its so-called  $R_j^*$  is lower:

$$R_j^* = (mK_j)/(r - m) \quad (\text{eq.1})$$

where  $m, K_j, r$  represent a species' mortality rate, half-saturation constant for uptake of  $R_j$ , and its maximum reproduction rate, respectively. The outcome of competition between X and Y can be graphically determined by plotting the isoclines of both species in the resource plane (solid lines in Fig. 1). An isocline is a line that connects the combinations of  $R_1$  and  $R_2$  at which

a species is at equilibrium, and its location is fully defined by the species'  $R_1^*$  and  $R_2^*$ . Thus, the two isoclines need to intersect for both species to have equilibrium densities at the same combination of resource levels. In the example of Fig. 1, there is a trade-off in the ability to take up nutrients: X is the best competitor for  $R_2$  ( $R_2^*$  of X <  $R_2^*$  of Y) while Y is the best competitor for  $R_1$  ( $R_1^*$  of Y <  $R_1^*$  of X). This trade-off allows the intersection of the two isoclines. However, this trade-off is a necessary but insufficient condition for coexistence: in addition, the resource supply point must lie within the coexistence region (in grey in Fig. 1), which is the region between the consumption vectors of both species (dashed lines in Fig. 1). Given the assumption of optimal foraging, the consumption vectors are proportional to  $R^*$ ; the per-capita consumption rate of a species on resource i is  $\frac{R_i^*}{y_i}$ , with  $y_i$  the amount of biomass produced per unit of resource i.

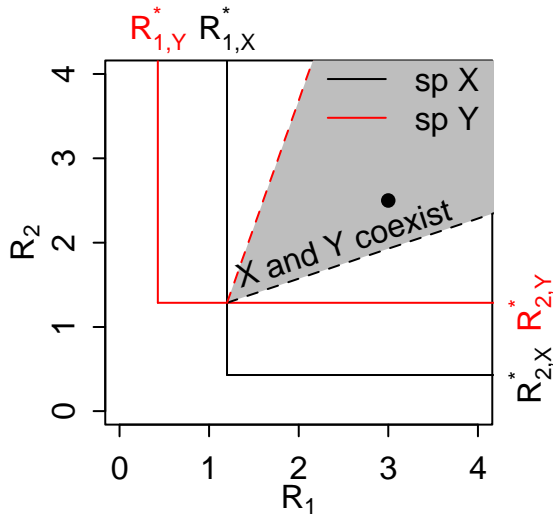


Figure 1: A plane of resource levels, containing two isoclines: one for species X (black), and one for species Y (red). Parameters  $(m, K_1, K_2, r)$  are  $(0.3, 2.8, 1, 1)$  for species X, and  $(0.3, 1, 3, 1)$  for species Y. Therefore,  $R_1^*$  for X,  $R_{1,X}^*=1.20$ ,  $R_{2,X}^*=0.43$ ,  $R_{1,Y}^*=0.43$ , and  $R_{2,Y}^*=1.29$ . The dashed lines are the two species' consumption vectors according to optimal foraging. The yield  $y$  is the same for both species (0.1). When the supply rate of both resources (dot) falls within the zone (in grey) delimited by the consumption vectors, coexistence is possible.

I consider a first pair of environmental change drivers that both slow down resource uptake: they both increase the half-saturation constant  $K_2$  of species X for  $R_2$ , and therefore  $R_2^*$  of X (eq.1), by a certain arbitrarily chosen factor (Fig. 2A-C). They do not affect any other parameters. A variety of mechanisms may underpin such effects. For example, temperature may alter nutrient-uptake proteins in plants (Giri *et al.*, 2017) and light-harvesting pigments in phytoplankton (Chalifour *et al.*, 2014), while ocean acidification can impair ciliary activity in mussels, decreasing feeding (Clements & Darrow, 2018). When present alone (Fig. 2A-B), both drivers do not affect species richness: the new coexistence region still comprises the resource supply point and so both species continue to coexist. However, when both drivers are present (Fig. 2C), the joint effect on species X's competitive ability is too large to maintain coexistence: the coexistence region does no longer contain the resource supply point. Hence, one species will go extinct. Thus, overall, the joint effect of these two drivers on species richness would be categorized as synergistic: the joint effect (a reduction from two to one species) is greater than expected from the single drivers, which have no effect on species richness when applied individually.

I now consider a second pair of drivers that work via the same mechanism as the first pair: both drivers again slow down resource uptake (Fig. 2D-F). However, in contrast to the first pair of drivers, both drivers now each target one specific species. This could be due to, for example, both species having different physiologies or resource uptake strategies, giving them different sensitivities to different stressors (Clements & Darrow, 2018; McMahon *et al.*, 2012; Mensens *et al.*, 2017). The first driver increases  $K_2$  of species X (Fig. 2D), while the second driver increases  $K_1$  of species Y (Fig. 2E). Again, they do not affect any other parameters. In both cases, the isoclines do not intersect anymore, so one species is excluded. However, when both drivers are present (Fig. 2F), the isoclines do intersect: the two drivers install a new trade-off



between the competitive abilities of species X and Y that saves them from extinction. Thus, the joint effect is smaller than expected based on the individual drivers. In a null model framework, this joint effect would be termed ‘antagonistic’. Taken together, the two pairs of drivers (Fig. 2A-C and Fig. 2D-F) illustrate that the same mechanism (a reduction of resource uptake) can lead to both synergistic and antagonistic effects.

I now turn to a third pair of drivers (Fig. 2G-I) that act on different mechanisms than the two first pairs given above. One driver (Fig. 2G) does slow down resource uptake (now it increases  $K_2$  of species Y) and leaves other parameters unchanged, as in the examples given before. However, the other driver corresponds to a decrease in the availability of  $R_1$  (Fig. 2H). The first driver causes extinction of one species by shifting the coexistence region sufficiently up such that it no longer includes the supply point (Fig. 2G). The second driver causes extinction too, by moving the resource supply point outside of the coexistence region (Fig. 2H). When combined, the joint effect of both drivers on richness is zero: both species are able to coexist because the new resource supply point falls into the new coexistence region (Fig. 2I). Thus, the null model approach would classify the joint effect as antagonistic. Taken together, the second and third pair of drivers illustrate that two different mechanisms (Fig. 2F: reductions of resource uptake vs. Fig. 2I: a reduction of resource uptake combined with a resource shift) can both lead to the same deviation from a null model prediction, in this case an antagonistic effect.

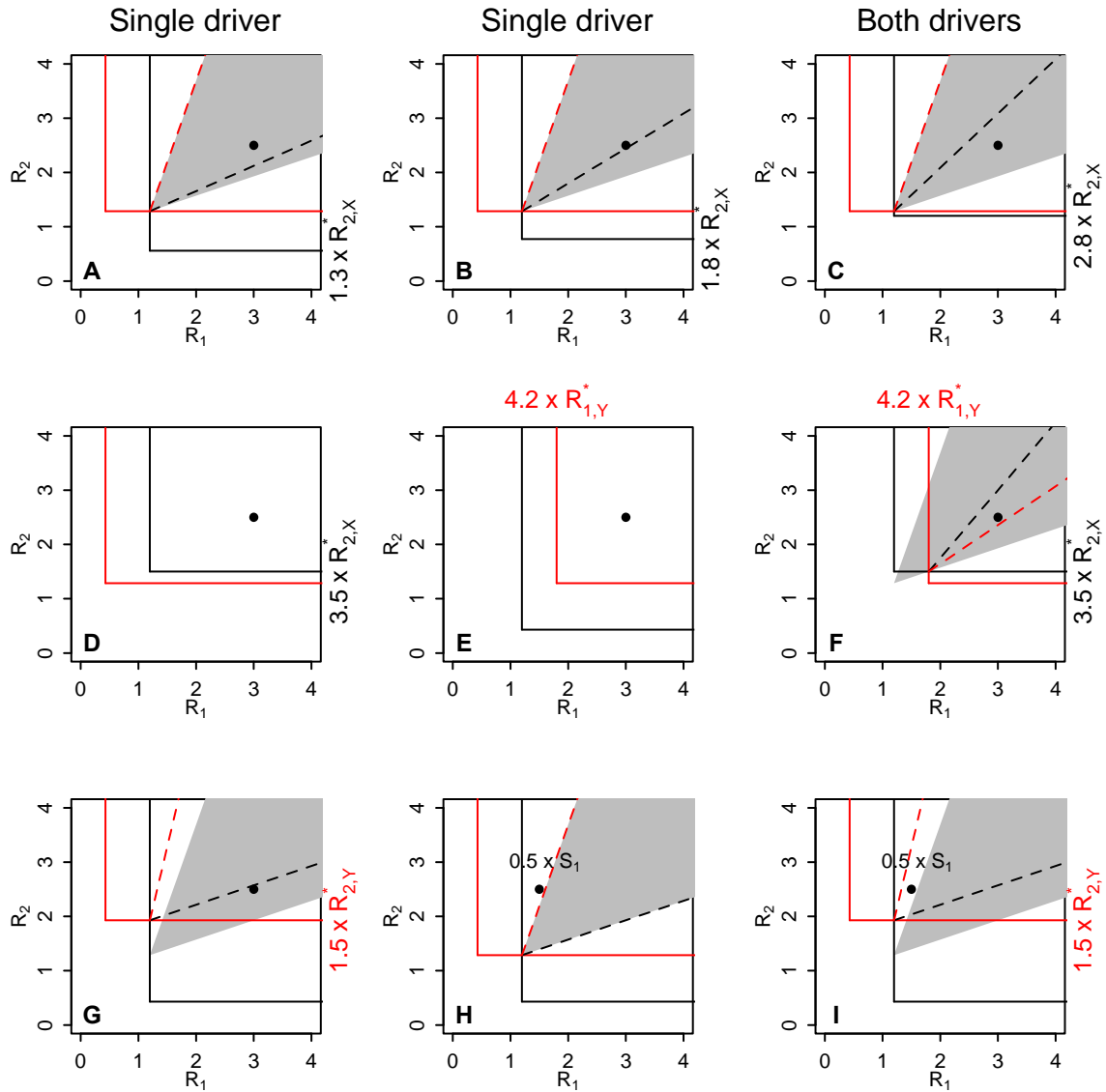


Figure 2: As Fig. 1, but now after exposure to one (panels A, B, D, E, G, H) or two environmental change drivers (columns C, F, I). Every row represents a different pair of drivers. The coexistence region in absence of change (in grey) is redrawn from Fig. 1 for comparison, unless when environmental change disrupts the intersection of isoclines and so removes the coexistence region (D, E). The numbers in black and red indicate the  $R^*$  values of species X and Y after modification by a stressor, respectively. The numbers inside panels H and I show the magnitude of resource supply shift.

One could argue that this example only shows that null models do not provide mechanistic insight, while they can still provide practical guidelines on how to manage ecosystems. Null models would allow grouping certain driver combinations that act, for example, synergistically. This information could be used for prioritisation and environmental decision making. However, the model illustration (Fig. 2) also shows that the deviations from null models will critically depend on the level of environmental change. For example, a smaller reduction of the resource uptake in Fig. 2G-I, or a larger reduction of  $R_1$ , would have resulted in different outcomes of how the joint effect is categorized. This illustrates the idea that information obtained from null models cannot be extrapolated beyond the tested ranges of the environmental change drivers (Schäfer & Piggott, 2018). This feature limits the capacity of null models to assist ecosystem management.

### **A new framework for community- and ecosystem-level effects of environmental changes**

As illustrated above, the current null model approach does not enhance understanding of community- and ecosystem-level effects of multiple drivers. We therefore need to move away from null model testing and start constructing a mechanistic framework (Griffen *et al.*, 2016; Kroeker *et al.*, 2017), as has been recently initiated at the level of populations (Galic *et al.*, 2017; Hodgson *et al.*, 2017) and communities (Thompson *et al.*, 2018). Here, I propose such a framework and show that deviations from its predictions can be informative, exactly because the assumptions underlying the predictions are well-defined.

As a starting point, I propose to formalise “environmental change”. I postulate that two main types of environmental change are resource supply shifts (Vitousek *et al.*, 1997) and the appearance of multiple abiotic stressors (Steinberg, 2012). The effects of resource supply shifts are a main ingredient of theoretical ecology, and so their effects on community and ecosystem

variables are well understood. Resource uptake theory (Armstrong & McGehee, 1976; Chase & Leibold, 2003; Harpole *et al.*, 2016; Hillebrand *et al.*, 2014; Huisman & Weissing, 1994; Tilman, 1982) predicts that an imbalance in resource supply, for example caused by changing nutrient loads, leads to biodiversity loss and altered ecosystem function (Loreau, 1998; Loreau, 2010) (e.g. as in Fig. 2H). Whether or not resource ratio shifts result in extinctions depends on the resource uptake traits of the species. Here, I define resource uptake traits (hereafter ‘traits’) as done by Litchman and Klausmeier (2008), i.e. as parameters of a resource uptake model. This definition is broadly applicable across a wide range of community types, including plankton (e.g. Edwards *et al.*, 2013), terrestrial plants (e.g. HilleRisLambers *et al.*, 2012) and animal consumers (Murdoch *et al.*, 2003), because resource uptake models are mathematically similar across community types. These traits mostly include half-saturation constants, conversion efficiencies from resource to biomass, and loss rates (Armstrong & McGehee, 1976; Harpole *et al.*, 2016; Hillebrand *et al.*, 2014; Huisman & Weissing, 1994; Tilman, 1982). These traits determine the location of the isoclines in Fig. 1 (e.g. half-saturation constants for nitrogen determine the  $R_N^*$ ), and therefore predict the outcome of competition at a given resource supply ratio. These traits thus determine how a species will respond to its biotic and abiotic environment and determine its contribution to ecosystem function, which makes them both response and effect traits (Violle *et al.*, 2007).

Inspired by the early work of Tilman *et al.* (1981), I postulate that integrating multiple stressors into resource uptake theory makes for a comprehensive framework to study community- and ecosystem-level effects of multiple drivers, including stressors and resource shifts. Specifically, I propose to realise this integration by making traits multivariate functions of multiple stressors (Fig. 3). If we know the effects of multiple stressors on such traits we can make educated guesses of how these effects translate to changes of variables at the level of

communities (e.g. biodiversity) and ecosystems (e.g. function). Note that the framework allows stressors to affect multiple traits, contrary to the examples provided in Fig. 2. In addition, we can readily include the additional influence of resource ratio shifts, since these are by definition covered by resource uptake theory. In this paper, I illustrate the framework for the case of competitive communities. However, resource uptake theory is general, and can be extended to food-webs (Chesson & Kuang, 2008; Murdoch *et al.*, 2003). Importantly, it can also be used to study community assembly (Koffel *et al.*, 2018; Seabloom *et al.*, 2003) and meta-community dynamics (Haegeman & Loreau, 2015; Tilman, 1994), allowing the inclusion of invasive species and habitat fragmentation as environmental change drivers not included in the present paper.

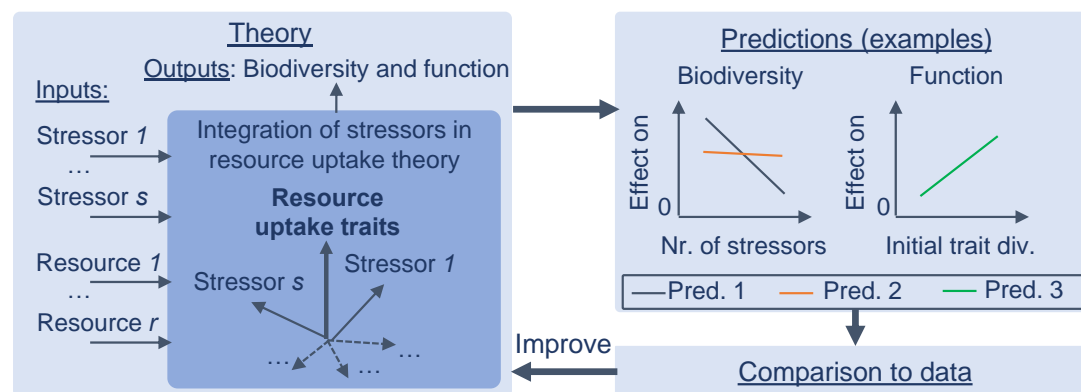


Figure 3: A proposed framework to study effects of multiple environmental changes on communities and ecosystems. Integrating multiple stressors into resource uptake theory yields predictions (pred.) regarding, for example, the influence of the number of stressors and the initial trait diversity of the exposed community on biodiversity and ecosystem function. Predictions 1-3 are detailed in the text. Comparing these predictions to experimental data can expose invalid assumptions underlying the theory, and therefore advances science.

## How the new framework advances comprehension

The proposed framework advances comprehension in three ways: (1) by formalising exposure to multiple stressors using a set of key factors; (2) by producing a multitude of testable predictions on how these factors, alone and in combination with resource ratio shifts, affect communities and ecosystems; and (3) by quantifying and interpreting deviations between these predictions and observed effects.

I identify five factors that can describe the exposure to a given stressor combination in a formal way. This formalisation facilitates mechanistic progress because these factors are key determinants of community- and ecosystem-level variables such as biodiversity and function. A first and second factor are evidently the number of stressors and stressor response intensity. In line with Steudel *et al.* (2012), I define stressor response intensity (*SRI*) as the community average (i.e. across all species) of the absolute effect of a particular stressor combination on population growth (e.g. biomass or number of individuals) in monoculture:

$$SRI = E[|M - M'|/M]$$

where  $E, M, M'$  represent the expected value operator, monoculture growth in absence of the stressor cocktail and in presence of that cocktail, respectively. Note that *SRI* could also be defined at the level of the traits, representing the community-average effect on traits, and could include information on the sign of the effect in monoculture (i.e. growth stimulation or depression). *SRI* depends on the tolerance of the exposed species and the value of the stressors and is therefore often referred to as ‘effect’ (e.g. Liess *et al.*, 2016; Schäfer & Piggott, 2018). *SRI* and the number of stressors are typically not considered separately in experimental studies (Brennan & Collins, 2015). However, scientific progress demands understanding the contribution of both factors to the net effect. For example, in Fig. 2C, the factor causing extinction is *SRI*, rather than the number of stressors. Indeed, exposing the community to only

one of the stressors, but at a higher *SRI*, could have sufficed to cause an extinction. A third factor is selectivity: the degree to which stressors affect all species or only a subset (De Laender *et al.*, 2016). For selective stressors, also co-tolerance plays a role: species can be (in)tolerant to multiple stressors at a time or not (Vinebrooke *et al.*, 2004). For example, in Fig. 2A-C, there is positive co-tolerance, while in Fig. 2D-F, there is negative co-tolerance, with radically different effects on species richness. This formalises the idea that selectivity is important only when it implies greater stress to species with particular traits (Diaz *et al.*, 2013; Newbold *et al.*, 2015; Suding *et al.*, 2008). Selective stressors can therefore cause different effects on biodiversity and function than unselective stressors (Mensens *et al.*, 2017; Selmants *et al.*, 2012; Spaak *et al.*, 2017). A fourth factor is initial trait diversity among species, which is expected to determine resistance to stressors. Initial trait diversity implying larger niche differences and smaller fitness differences will be better buffered (i.e. more resistant) to stressor effects (Chesson, 2000). A fifth factor is the dynamic behaviour of stressors. Indeed, stressors often fluctuate through time (Gunderson *et al.*, 2016), which can cause different effects on coexistence (Chesson, 1994), and thus on biodiversity and function, than when stressors remain constant through time.

Defining the five factors allows representing stressor combinations in a standardized way. However, more importantly, we can make general predictions on how these factors matter for the resulting community- and ecosystem-level effects (Fig. 3), and on how these factors combine with resource ratio shifts. These predictions can offer scientists, working on a variety of communities, theory-based and testable predictions that extend beyond the question if effects are, for example, additive. More specifically, for various combinations of the number of stressors, *SRI*, selectivity and co-tolerance, initial trait diversity, and temporal stressor dynamics (constant, or various kinds of fluctuations, as in Jiang & Morin, 2004; Jiang & Morin,

2007), various aspects of biodiversity but also two types of ecosystem function (biomass production and resource uptake) can be predicted. Here, as an example, I provide five such predictions, of which three are illustrated in Fig. 3. The exhaustive analysis of how the five factors mentioned above together affect biodiversity and function is not an objective of the present opinion paper. Future theoretical contributions can address this question in full-factorial setups. Rather, I want to illustrate the kind of predictions the framework could produce. Afterwards, I discuss how these could be experimentally tested.

Prediction 1 postulates that, for selective stressors exhibiting negative co-tolerance (each stressor only affects one species), the effect on species richness decreases with the number of stressors. This is because, with an increasing number of stressors, but a constant *SRI*, an increasing number of species will be affected but to a smaller extent. When *SRI* is low, this will not affect the trade-offs that maintain composition and sustain function. When *SRI* is high, this can lead to new trade-offs being installed (e.g. Fig. 2F). Note that, for both low and high *SRI*, effects on biomass production can occur, leading to stressor effects on function in absence of effects on richness (Spaak *et al.*, 2017). Prediction 2 postulates that lower stressor selectivity results in smaller effects of the number of stressors on biodiversity and function. For example, when stressors are completely unselective, every stressor should affect every species to exactly the same extent. The number of stressors causing this effect is irrelevant. In the opposite case, when stressors are very selective, i.e. every stressor affects a different species, the number of affected species equals the number of stressors. A higher number of stressors thus means that more species are affected, but to a lesser extent (again, since *SRI* needs to stay constant). Prediction 3 postulates that the effects of selective stressors on function increase with initial trait diversity. For example, when initial trait diversity is small (all species have very similar trait values), small levels of selective stress can suffice to disrupt coexistence, leading to



diversity loss. However, since tolerant and sensitive species have very similar trait values, tolerant species will rapidly compensate for the lost biomass of the sensitive species and changes of function will be minor. When initial trait diversity is high, selective stress will be less likely to cause exclusion but effects on function can be more substantial because of a loss of functional complementarity. Prediction 4 postulates that fluctuations of stressors matter less when the number of stressors is low. This is because, when resource supply does not fluctuate, affecting few types (a consequence of the current example of high stressor selectivity) is less likely to cause large changes in resource levels. This reduces the covariation between environmental and competition effects on species' growth. Coexistence theory learns that this makes fluctuation-dependent coexistence mechanisms less important (Chesson, 1994). Finally, Fig. 2I illustrates prediction 5: stressors that reduce resource uptake can make community composition less resistant to resource ratio shifts. Indeed, the two stressors reduced the coexistence area, meaning that smaller shifts of resource ratio are needed to disrupt coexistence and cause extinction.

I propose testing predictions like those listed above using two approaches. A first approach could focus on 'high-level' predictions that transcend the specificities of model systems. Such high-level predictions exist in other research fields. Examples include the increase of ecosystem function with the number of functionally different species (Cardinale *et al.*, 2011), the effects of niche dimensionality on species diversity (Harpole *et al.*, 2016), and the effect of dispersal on regional biodiversity (Mouquet & Loreau, 2003). The proposed framework could be exploited to identify predictions about the effects of the number of stressors, *SRI*, and selectivity that are robust to other parameter settings. Monoculture experiments would then suffice to quantify *SRI* and selectivity for a given community composition exposed to a given stressor combination: resource uptake traits need not be measured. Thus, by selecting the right

combination of species and stressors, one can experimentally test high-level predictions on how these factors, alone and combined with resource ratio shifts, affect biodiversity and function.

A second approach is far more labour intensive and requires growing species in monoculture, at various resource supply and stressor levels, to infer their traits and how these change with stress. These data can then predict the outcome of competition at various combinations of stressors and resource ratio shifts. Comparing these predictions to the actual outcome of competition experiments is the strongest possible test of the theory. This approach is conceptually similar to the work by Tilman *et al.* (1981) for the case of a single stressor, temperature. However, the amount of work needed to apply this approach to multiple stressors restricts this approach to microbial systems tested using high-throughput protocols (Altermatt *et al.*, 2015), and makes it inapplicable to species with longer life cycles. Still, this approach can lead to considerable progress in two ways. First, it would yield insight into which facets of global change most disturb trade-offs that sustain biodiversity and functions in microbial systems (Litchman *et al.*, 2015). Second, it would add environmental stress as a new dimension to the classic topic of resource enrichment and niche dimensionality (Borer *et al.*, 2014; Cardinale *et al.*, 2009; Harpole *et al.*, 2016; Hillebrand *et al.*, 2014).

One useful ingredient of the existing null model approach is the fact that it makes clear predictions. Assessing the capacity and limitations of predictive capacity is indeed a central goal in ecology and environmental science (Houlahan *et al.*, 2017; Petchey *et al.*, 2015). However, the new framework I propose here allows studying not only predictive capacity but also what drives predictive capacity, again because the assumptions are clear and are different from the predictions. Thus, observations deviating from theoretical predictions (e.g.

predictions 1-5 listed above) point at invalid assumptions and thus expose areas of theory improvement, advancing science. For example, the theoretical approach I propose assumes that the interaction type does not change with *SRI*: resource competition is always the only interaction type. The only way stressors can change community dynamics is by changing resource uptake traits, and thus potentially competitive outcome. Regardless of the *SRI*, species always interact indirectly by relying on the same resources; they never interact directly. Thus, when species interaction types change profoundly with *SRI* we can expect theoretical predictions to become increasingly erroneous (Barton & Ives, 2014; Mulder *et al.*, 2001; Suttle *et al.*, 2007). More specifically, the stress gradient hypothesis postulates that competitive (i.e. negative) interactions would switch to facilitative (i.e. positive) interactions (Olsen *et al.*, 2016) as environmental change intensifies (i.e. the environment gets ‘harsher’, or *SRI* increases) (He & Bertness, 2014; Hines *et al.*, 2015; Maestre *et al.*, 2009; Mulder *et al.*, 2001). Thus, in community types where predictive capacity drops with *SRI*, more detailed studies could be set up to examine interaction types along stressor gradients. In an experimental study, Vanellander *et al.* (2009) found that the marine diatom *Cylindrotheca closterium* grew better in medium containing substances leaked by other diatom species than in fresh medium. In a modelling study, Baert *et al.* (2016) inferred from experimental data that such facilitative interactions could be the exception rather than the rule in a similar community type evaluated along a pesticide gradient.

Another example of how the proposed theory could deviate from observed joint effects is through the influence of time. Again, the theory is very clear on how time is considered: it assumes traits to change instantly with the stressor level (Fig. 3). Community dynamics are therefore purely driven by external fluctuations in *SRI*, by the rate at which ecological interactions propagate these fluctuations to population growth, and by consumer-resource

dynamics internal to the system (Chesson, 1994; Huisman & Weissing, 1999). Thus, one can expect a reduced predictive capacity when long-term effects on traits occur, e.g. due to evolution or slow transgenerational plasticity (Doebeli & Ispolatov, 2014; Litchman *et al.*, 2015). This is because, in this case, the resulting long-term effects on biodiversity and function will be unpredictable from short-term responses of traits to environmental change (Feckler *et al.*, 2018; Turcotte & Levine, 2017). Microbial systems are well-suited to study such effects in realistic time frames, as one can study adaptation of single strains to new environmental conditions or altered fluctuation regimes, due to novel mutations or horizontal gene transfer (Brennan & Collins, 2015; Collins *et al.*, 2014; Litchman *et al.*, 2015). For phytoplankton, such events have been found to be more prevalent when the number of environmental drivers is greater (Brennan *et al.*, 2017). Thus, when the predictive capacity of the proposed theory (Fig. 3) decreases with the number of generations and does so more when the number of drivers is higher, this could stimulate studies on, for example, the adaptation of resource uptake traits.

Both examples given here of how the proposed theory could misjudge the effects of the number and kind of drivers on communities and ecosystems align with the idea that science benefits from ‘breaking models’ when these models have formal assumptions (Thiele & Grimm, 2015). Indeed, this practice not only identifies follow-up studies on facilitation and trait adaptation but will also lead to extensions of the theory. Resource uptake theory can be extended with equations for exudate production and uptake (Van den Meersche *et al.*, 2004), and for long-term trait change. As a starting point for long-term trait change, one could consider approaches from quantitative genetics, allowing to make various assumptions on the direction and rate of trait adaptation along gradients of environmental change (Barabás & D'Andrea, 2016; Chevin *et al.*, 2010; Schreiber *et al.*, 2011).

## Conclusions

In order to advance understanding of how multiple drivers of environmental change affect communities and ecosystems, we need to move away from testing null models: they lack a mechanistic basis and deviations from, or correspondence to, observations can therefore not advance insight. Here, I propose to base predictions on ecological theory and interpret deviations from observations as an essential step to meet the scientific and societal challenge of understanding global change effects. The approach I advocate here can probably be generalized to other organizational levels, because quantitative frameworks are available to predict physiological effects of multiple stressors at the individual and population level, including bio-energetic models (Sokolova, 2013) and dynamic energy budget models coupled with individual-based models (Galic *et al.*, 2017; Goussen *et al.*, 2016).

At present, there is a tendency to label deviations from additivity as ‘ecological surprises’ (Brook *et al.*, 2008; Crain *et al.*, 2008; Darling & Cote, 2008; Thompson *et al.*, 2018), implying they are not to be expected (‘surprising’) on the basis of ecological science. However, synergistic and antagonistic effects can be expected from basic ecological science and are therefore no ecological surprises (Fig. 2 and Halstead *et al.*, 2014). The proposed framework provides better guidance as to which driver combinations provoke truly surprising effects, i.e. effects that are not expected from one of ecology’s most established theories but triggered by emergent processes such as stress-induced facilitation and trait adaptation.

## Figure captions

Embedded in the text.

## Figures

Embedded in the text.

## References

- Altermatt, F., Fronhofer, E. A., Garnier, A., Giometto, A., Hammes, F., Klecka, J., Legrand, D., Mächler, E., Massie, T. M., Pennekamp, F., Plebani, M., Pontarp, M., Schtickzelle, N., Thuillier, V., & Petchey, O. L. (2015). Big answers from small worlds: a user's guide for protist microcosms as a model system in ecology and evolution. *Methods in Ecology and Evolution*, **6**, 218-231.
- Armstrong, R. A., & McGehee, R. (1976). Coexistence of species competing for shared resources. *Theoretical Population Biology*, **9**, 317-328.
- Baert, J. M., Janssen, C. R., Sabbe, K., & De Laender, F. (2016). Per capita interactions and stress tolerance drive stress-induced changes in biodiversity effects on ecosystem functions. *Nature Communications*, **7**, 12486.
- Baert, J. M., Jaspers, S., Janssen, C. R., Laender, F. D., Aerts, M., & Chisholm, R. (2017). Nonlinear partitioning of biodiversity effects on ecosystem functioning. *Methods in Ecology and Evolution*, **8**, 1233-1240.
- Barabás, G., & D'Andrea, R. (2016). The effect of intraspecific variation and heritability on community pattern and robustness. *Ecology Letters*, **19**, 977-986.
- Barton, B. T., & Ives, A. R. (2014). Species interactions and a chain of indirect effects driven by reduced precipitation. *Ecology*, **95**, 486-494.
- Borer, E. T., Seabloom, E. W., Gruner, D. S., Harpole, W. S., Hillebrand, H., Lind, E. M., Adler, P. B., Alberti, J., Anderson, T. M., Bakker, J. D., Biederman, L., Blumenthal, D., Brown, C. S., Brudvig, L. A., Buckley, Y. M., Cadotte, M., Chu, C., Cleland, E. E., Crawley, M. J., Daleo, P., Damschen, E. I., Davies, K. F., DeCrappeo, N. M., Du, G., Firn, J., Hautier, Y., Heckman, R. W., Hector, A., HilleRisLambers, J., Iribarne, O., Klein, J. A., Knops, J. M. H., La Pierre, K. J., Leakey, A. D. B., Li, W., MacDougall, A. S., McCulley, R. L., Melbourne, B. A., Mitchell, C. E., Moore, J. L., Mortensen, B.,

496 O'Halloran, L. R., Orrock, J. L., Pascual, J., Prober, S. M., Pyke, D. A., Risch, A. C.,  
 497 Schuetz, M., Smith, M. D., Stevens, C. J., Sullivan, L. L., Williams, R. J., Wragg, P.  
 498 D., Wright, J. P., & Yang, L. H. (2014). Herbivores and nutrients control grassland  
 499 plant diversity via light limitation. *Nature*, **508**, 517.

500 Brennan, G., & Collins, S. (2015). Growth responses of a green alga to multiple environmental  
 501 drivers. *Nature Climate Change*, **5**, 892.

502 Brennan, G. L., Colegrave, N., & Collins, S. (2017). Evolutionary consequences of multidriver  
 503 environmental change in an aquatic primary producer. *Proceedings of the National*  
 504 *Academy of Sciences*, **114**, 9930-9935.

505 Brook, B. W., Sodhi, N. S., & Bradshaw, C. J. A. (2008). Synergies among extinction drivers  
 506 under global change. *Trends in Ecology & Evolution*, **23**, 453-460.

507 Cardinale, B. J., Hillebrand, H., Harpole, W. S., Gross, K., & Ptacnik, R. (2009). Separating  
 508 the influence of resource 'availability' from resource 'imbalance' on productivity–  
 509 diversity relationships. *Ecology Letters*, **12**, 475-487.

510 Cardinale, B. J., Matulich, K. L., Hooper, D. U., Byrnes, J. E., Duffy, E., Gamfeldt, L.,  
 511 Balvanera, P., O'Connor, M. I., & Gonzalez, A. (2011). The functional role of producer  
 512 diversity in ecosystems. *American Journal of Botany*, **98**, 572-592.

513 Chalifour, A., Arts, M. T., Kainz, M. J., & Juneau, P. (2014). Combined effect of temperature  
 514 and bleaching herbicides on photosynthesis, pigment and fatty acid composition of  
 515 *Chlamydomonas reinhardtii*. *European Journal of Phycology*, **49**, 508-515.

516 Chase, J. M., & Leibold, M. (2003). *Ecological Niches: Linking Classical and Contemporary*  
 517 *Approaches*. Chicago: University of Chicago Press.

518 Chesson, P. (1994). Multispecies Competition in Variable Environments. *Theoretical*  
 519 *Population Biology*, **45**, 227-276.

520 Chesson, P. (2000). Mechanisms of maintenance of species diversity. *Annual Review of*  
521 *Ecology, Evolution, and Systematics*, **31**, 343–366.

522 Chesson, P., & Kuang, J. J. (2008). The interaction between predation and competition. *Nature*,  
523 **456**, 235-238.

524 Chevin, L.-M., Lande, R., & Mace, G. M. (2010). Adaptation, Plasticity, and Extinction in a  
525 Changing Environment: Towards a Predictive Theory. *PLoS Biology*, **8**, e1000357.

526 Clements, J. C., & Darrow, E. S. (2018). Eating in an acidifying ocean: a quantitative review  
527 of elevated CO<sub>2</sub> effects on the feeding rates of calcifying marine invertebrates.  
528 *Hydrobiologia*.

529 Collins, S., Rost, B., & Rynearson, T. A. (2014). Evolutionary potential of marine  
530 phytoplankton under ocean acidification. *Evolutionary Applications*, **7**, 140-155.

531 Côté, I. M., Darling, E. S., & Brown, C. J. (2016). Interactions among ecosystem stressors and  
532 their importance in conservation. *Proceedings of the Royal Society of London B:*  
533 *Biological Sciences*, **283**.

534 Crain, C. M., Kroeker, K., & Halpern, B. S. (2008). Interactive and cumulative effects of  
535 multiple human stressors in marine systems. *Ecology Letters*, **11**, 1304-1315.

536 Darling, E. S., & Cote, I. M. (2008). Quantifying the evidence for ecological synergies.  
537 *Ecology Letters*, **11**, 1278-1286.

538 De Laender, F., Rohr, J. R., Ashauer, R., Baird, D. J., Berger, U., Eisenhauer, N., Grimm, V.,  
539 Hommen, U., Maltby, L., Melià, C. J., Pomati, F., Roessink, I., Radchuk, V., & Van  
540 den Brink, P. J. (2016). Reintroducing Environmental Change Drivers in Biodiversity-  
541 Ecosystem Functioning Research. *Trends in Ecology & Evolution*, **31**, 905-915.

542 Diaz, S., Purvis, A., Cornelissen, J. H., Mace, G. M., Donoghue, M. J., Ewers, R. M., Jordano,  
543 P., & Pearse, W. D. (2013). Functional traits, the phylogeny of function, and ecosystem  
544 service vulnerability. *Ecology and Evolution*, **3**, 2958-2975.



545 Doebeli, M., & Ispolatov, I. (2014). Chaos and unpredictability in evolution. *Evolution*, **68**,  
546 1365-1373.

547 Dupont, S., & Pörtner, H. (2013). Get ready for ocean acidification. *Nature*, **498**, 429.

548 Edwards, K. F., Litchman, E., & Klausmeier, C. A. (2013). Functional traits explain  
549 phytoplankton community structure and seasonal dynamics in a marine ecosystem.  
550 *Ecology Letters*, **16**, 56-63.

551 Feckler, A., Goedkoop, W., Konschak, M., Bundschuh, R., Kenngott, K. G. J., Schulz, R.,  
552 Zubrod, J. P., & Bundschuh, M. (2018). History matters: Heterotrophic microbial  
553 community structure and function adapt to multiple stressors. *Global Change Biology*,  
554 **24**, e402-e415.

555 Fox, J. W. (2005). Interpreting the selection effect of biodiversity on ecosystem function.  
556 *Ecology Letters*, **8**, 846-856.

557 Galic, N., Grimm, V., & Forbes, V. E. (2017). Impaired ecosystem process despite little effects  
558 on populations: modeling combined effects of warming and toxicants. *Global Change*  
559 *Biology*, **23**, 2973-2989.

560 Garnier, A., Pennekamp, F., Lemoine, M., & Petchey, O. L. (2017). Temporal scale dependent  
561 interactions between multiple environmental disturbances in microcosm ecosystems.  
562 *Glob Chang Biol*, **23**, 5237-5248.

563 Giri, A., Heckathorn, S., Mishra, S., & Krause, C. (2017). Heat Stress Decreases Levels of  
564 Nutrient-Uptake and -Assimilation Proteins in Tomato Roots. *Plants*, **6**, 6.

565 Goussen, B., Price, O. R., Rendal, C., & Ashauer, R. (2016). Integrated presentation of  
566 ecological risk from multiple stressors. *Scientific Reports*, **6**, 36004.

567 Griffen, B. D., Belgrad, B. A., Cannizzo, Z. J., Knotts, E. R., & Hancock, E. R. (2016).  
568 Rethinking our approach to multiple stressor studies in marine environments. *Marine*  
569 *Ecology Progress Series*, **543**, 273-281.

570 Gunderson, A. R., Armstrong, E. J., & Stillman, J. H. (2016). Multiple Stressors in a Changing  
571 World: The Need for an Improved Perspective on Physiological Responses to the  
572 Dynamic Marine Environment. *Annual Review of Marine Science*, **8**, 357-378.

573 Häder, D.-P., & Gao, K. (2015). Interactions of anthropogenic stress factors on marine  
574 phytoplankton. *Frontiers in Environmental Science*, **3**.

575 Haegeman, B., & Loreau, M. (2015). A Graphical-Mechanistic Approach to Spatial Resource  
576 Competition. *The American Naturalist*, **185**, E1-E13.

577 Halpern, B. S., Frazier, M., Potapenko, J., Casey, K. S., Koenig, K., Longo, C., Lowndes, J.  
578 S., Rockwood, R. C., Selig, E. R., Selkoe, K. A., & Walbridge, S. (2015). Spatial and  
579 temporal changes in cumulative human impacts on the world's ocean. *Nature*  
580 *Communications*, **6**, 7615.

581 Halpern, B. S., Walbridge, S., Selkoe, K. A., Kappel, C. V., Micheli, F., D'Agrosa, C., Bruno,  
582 J. F., Casey, K. S., Ebert, C., Fox, H. E., Fujita, R., Heinemann, D., Lenihan, H. S.,  
583 Madin, E. M. P., Perry, M. T., Selig, E. R., Spalding, M., Steneck, R., & Watson, R.  
584 (2008). A Global Map of Human Impact on Marine Ecosystems. *Science*, **319**, 948-  
585 952.

586 Halstead, N. T., McMahon, T. A., Johnson, S. A., Raffel, T. R., Romansic, J. M., Crumrine, P.  
587 W., & Rohr, J. R. (2014). Community ecology theory predicts the effects of  
588 agrochemical mixtures on aquatic biodiversity and ecosystem properties. *Ecology*  
589 *Letters*, **17**, 932-941.

590 Harpole, W. S., Sullivan, L. L., Lind, E. M., Firn, J., Adler, P. B., Borer, E. T., Chase, J., Fay,  
591 P. A., Hautier, Y., Hillebrand, H., MacDougall, A. S., Seabloom, E. W., Williams, R.,  
592 Bakker, J. D., Cadotte, M. W., Chaneton, E. J., Chu, C., Cleland, E. E., D'Antonio, C.,  
593 Davies, K. F., Gruner, D. S., Hagenah, N., Kirkman, K., Knops, J. M. H., La Pierre, K.  
594 J., McCulley, R. L., Moore, J. L., Morgan, J. W., Prober, S. M., Risch, A. C., Schuetz,

595 M., Stevens, C. J., & Wragg, P. D. (2016). Addition of multiple limiting resources  
 596 reduces grassland diversity. *Nature*, **537**, 93.

597 He, Q., & Bertness, M. D. (2014). Extreme stresses, niches, and positive species interactions  
 598 along stress gradients. *Ecology*, **95**, 1437-1443.

599 Hillebrand, H., Cowles, J. M., Lewandowska, A., Van de Waal, D. B., & Plum, C. (2014).  
 600 Think ratio! A stoichiometric view on biodiversity–ecosystem functioning research.  
 601 *Basic and Applied Ecology*, **15**, 465-474.

602 HilleRisLambers, J., Adler, P. B., Harpole, W. S., Levine, J. M., & Mayfield, M. M. (2012).  
 603 Rethinking Community Assembly through the Lens of Coexistence Theory. *Annual*  
 604 *Review of Ecology, Evolution, and Systematics*, **43**, 227-248.

605 Hines, J., Eisenhauer, N., & Drake, B. G. (2015). Inter-annual changes in detritus-based food  
 606 chains can enhance plant growth response to elevated atmospheric CO<sub>2</sub>. *Global*  
 607 *Change Biology*, **21**, 4642-4650.

608 Hodgson, E. E., Essington, T. E., & Halpern, B. S. (2017). Density dependence governs when  
 609 population responses to multiple stressors are magnified or mitigated. *Ecology*, **98**,  
 610 2673-2683.

611 Houlahan, J. E., McKinney, S. T., Anderson, T. M., & McGill, B. J. (2017). The priority of  
 612 prediction in ecological understanding. *Oikos*, **126**, 1-7.

613 Huisman, J., & Weissing, F. J. (1994). Light-Limited Growth and Competition for Light in  
 614 Well-Mixed Aquatic Environments: An Elementary Model. *Ecology*, **75**, 507-520.

615 Huisman, J., & Weissing, F. J. (1999). Biodiversity of plankton by species oscillations and  
 616 chaos. *Nature*, **402**, 407.

617 Jackson, M. C., Loewen, C. J. G., Vinebrooke, R. D., & Chimimba, C. T. (2016). Net effects  
 618 of multiple stressors in freshwater ecosystems: a meta-analysis. *Global Change*  
 619 *Biology*, **22**, 180-189.

620 Jiang, L., & Morin, P. J. (2004). Temperature-dependent interactions explain unexpected  
621 responses to environmental warming in communities of competitors. *Journal of Animal*  
622 *Ecology*, **73**, 569-576.

623 Jiang, L. I. N., & Morin, P. J. (2007). Temperature fluctuation facilitates coexistence of  
624 competing species in experimental microbial communities. *Journal of Animal Ecology*,  
625 **76**, 660-668.

626 Koffel, T., Boudsocq, S., Loeuille, N., & Daufresne, T. (2018). Facilitation- vs. competition-  
627 driven succession: the key role of resource-ratio. *Ecology Letters*, **0**.

628 Kroeker, K. J., Kordas, R. L., & Harley, C. D. G. (2017). Embracing interactions in ocean  
629 acidification research: confronting multiple stressor scenarios and context dependence.  
630 *Biology Letters*, **13**.

631 Liess, M., Foit, K., Knillmann, S., Schäfer, R. B., & Liess, H.-D. (2016). Predicting the synergy  
632 of multiple stress effects. *Scientific Reports*, **6**, 32965.

633 Litchman, E., Edwards, K. F., & Klausmeier, C. A. (2015). Microbial resource utilization traits  
634 and trade-offs: implications for community structure, functioning, and biogeochemical  
635 impacts at present and in the future. *Frontiers in Microbiology*, **6**.

636 Litchman, E., & Klausmeier, C. A. (2008). Trait-Based Community Ecology of Phytoplankton.  
637 *Annual Review of Ecology, Evolution, and Systematics*, **39**, 615-639.

638 Loreau, M. (1998). Biodiversity and ecosystem functioning: A mechanistic model.  
639 *Proceedings of the National Academy of Sciences of the United States of America*, **95**,  
640 5632-5636.

641 Loreau, M. (2010). Linking biodiversity and ecosystems: towards a unifying ecological theory.  
642 *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, **365**,  
643 49-60.

644 Loreau, M., & Hector, A. (2001). Partitioning selection and complementarity in biodiversity  
645 experiments. *Nature*, **412**, 72.

646 Maestre, F. T., Callaway, R. M., Valladares, F., & Lortie, C. J. (2009). Refining the stress-  
647 gradient hypothesis for competition and facilitation in plant communities. *Journal of*  
648 *Ecology*, **97**, 199-205.

649 McMahon, T. A., Halstead, N. T., Johnson, S., Raffel, T. R., Romansic, J. M., Crumrine, P.  
650 W., & Rohr, J. R. (2012). Fungicide-induced declines of freshwater biodiversity modify  
651 ecosystem functions and services. *Ecology Letters*, **15**, 714-722.

652 Mensens, C., De Laender, F., Janssen, C. R., Sabbe, K., & De Troch, M. (2017). Different  
653 response–effect trait relationships underlie contrasting responses to two chemical  
654 stressors. *Journal of Ecology*, **105**, 1598–1609.

655 Mouquet, N., & Loreau, M. (2003). Community patterns in source-sink metacommunities. *Am*  
656 *Nat*, **162**, 544-557.

657 Mulder, C. P. H., Uliassi, D. D., & Doak, D. F. (2001). Physical stress and diversity-  
658 productivity relationships: The role of positive interactions. *Proceedings of the*  
659 *National Academy of Sciences of the United States of America*, **98**, 6704-6708.

660 Murdoch, W. M., Briggs, C. J., & Nisbet, R. M. (2003). *Consumer-resource dynamic* (Vol.  
661 36). Princeton and Oxford: Princeton University Press.

662 Newbold, T., Hudson, L. N., Hill, S. L. L., Contu, S., Lysenko, I., Senior, R. A., Borger, L.,  
663 Bennett, D. J., Choimes, A., Collen, B., Day, J., De Palma, A., Diaz, S., Echeverria-  
664 Londono, S., Edgar, M. J., Feldman, A., Garon, M., Harrison, M. L. K., Alhusseini, T.,  
665 Ingram, D. J., Itescu, Y., Kattge, J., Kemp, V., Kirkpatrick, L., Kleyer, M., Correia, D.  
666 L. P., Martin, C. D., Meiri, S., Novosolov, M., Pan, Y., Phillips, H. R. P., Purves, D.  
667 W., Robinson, A., Simpson, J., Tuck, S. L., Weiher, E., White, H. J., Ewers, R. M.,

668 Mace, G. M., Scharlemann, J. P. W., & Purvis, A. (2015). Global effects of land use on  
669 local terrestrial biodiversity. *Nature*, **520**, 45-50.

670 Olsen, S. L., Töpper, J. P., Skarpaas, O., Vandvik, V., & Klanderud, K. (2016). From  
671 facilitation to competition: temperature-driven shift in dominant plant interactions  
672 affects population dynamics in seminatural grasslands. *Global Change Biology*, **22**,  
673 1915-1926.

674 Petchey, O. L., Pontarp, M., Massie, T. M., Kéfi, S., Ozgul, A., Weilenmann, M., Palamara,  
675 G. M., Altermatt, F., Matthews, B., Levine, J. M., Childs, D. Z., McGill, B. J.,  
676 Schaepman, M. E., Schmid, B., Spaak, P., Beckerman, A. P., Pennekamp, F., & Pearse,  
677 I. S. (2015). The ecological forecast horizon, and examples of its uses and determinants.  
678 *Ecology Letters*, **18**, 597-611.

679 Piggott, J. J., Townsend, C. R., & Matthaei, C. D. (2015). Reconceptualizing synergism and  
680 antagonism among multiple stressors. *Ecology and Evolution*, **5**, 1538-1547.

681 Rudd, M. A. (2014). Scientists' perspectives on global ocean research priorities. *Frontiers in*  
682 *Marine Science*, **1**.

683 Schäfer, R. B., Kühn, B., Malaj, E., König, A., & Gergs, R. (2016). Contribution of organic  
684 toxicants to multiple stress in river ecosystems. *Freshwater Biology*, **61**, 2116-2128.

685 Schäfer, R. B., & Piggott, J. J. (2018). Advancing understanding and prediction in multiple  
686 stressor research through a mechanistic basis for null models. *Global Change Biology*.

687 Schreiber, S. J., Bürger, R., & Bolnick, D. I. (2011). The community effects of phenotypic and  
688 genetic variation within a predator population. *Ecology*, **92**, 1582-1593.

689 Seabloom, E. W., Harpole, W. S., Reichman, O. J., & Tilman, D. (2003). Invasion, competitive  
690 dominance, and resource use by exotic and native California grassland species. *Proc*  
691 *Natl Acad Sci U S A*, **100**, 13384-13389.

692 Selmants, P. C., Zavaleta, E. S., Pasari, J. R., & Hernandez, D. L. (2012). Realistic plant species  
693 losses reduce invasion resistance in a California serpentine grassland. *Journal of*  
694 *Ecology*, **100**, 723-731.

695 Sokolova, I. M. (2013). Energy-limited tolerance to stress as a conceptual framework to  
696 integrate the effects of multiple stressors. *Integr Comp Biol*, **53**, 597-608.

697 Spaak, J. W., Baert, J. M., Baird, D. J., Eisenhauer, N., Maltby, L., Pomati, F., Radchuk, V.,  
698 Rohr, J. R., Van den Brink, P. J., & De Laender, F. (2017). Shifts of community  
699 composition and population density substantially affect ecosystem function despite  
700 invariant richness. *Ecology Letters*, **20**, 1315-1324.

701 Steinberg, C. E. W. (2012). *Stress Ecology. Environmental Stress as Ecological Driving Force*  
702 *and Key Player in Evolution.*: Springer.

703 Steudel, B., Hector, A., Friedl, T., Lofke, C., Lorenz, M., Wesche, M., Kessler, M., & Gessner,  
704 M. (2012). Biodiversity effects on ecosystem functioning change along environmental  
705 stress gradients. *Ecology Letters*, **15**, 1397-1405.

706 Suding, K. N., Lavorel, S., Chapin, F. S., Cornelissen, J. H. C., DÍAz, S., Garnier, E., Goldberg,  
707 D., Hooper, D. U., Jackson, S. T., & Navas, M.-L. (2008). Scaling environmental  
708 change through the community-level: a trait-based response-and-effect framework for  
709 plants. *Global Change Biology*, **14**, 1125-1140.

710 Suttle, K. B., Thomsen, M. A., & Power, M. E. (2007). Species Interactions Reverse Grassland  
711 Responses to Changing Climate. *Science*, **315**, 640-642.

712 Thiele, J. C., & Grimm, V. (2015). Replicating and breaking models: good for you and good  
713 for ecology. *Oikos*, **124**, 691-696.

714 Thompson, P. L., MacLennan, M. M., & Vinebrooke, R. D. (2018). An improved null model  
715 for assessing the net effects of multiple stressors on communities. *Global Change*  
716 *Biology*, **24**, 517-525.

717 Tilman, D. (1982). *Resource Competition and Community Structure* (Vol. 17): Princeton.

718 Tilman, D. (1994). Competition and Biodiversity in Spatially Structured Habitats. *Ecology*, **75**,  
719 2-16.

720 Tilman, D., Mattson, M., & Langer, S. (1981). Competition and nutrient kinetics along a  
721 temperature gradient: An experimental test of a mechanistic approach to niche theory.  
722 *Limnology and Oceanography*, **26**, 1020-1033.

723 Tolkkinen, M., Mykra, H., Annala, M., Markkola, A. M., Vuori, K. M., & Muotka, T. (2015).  
724 Multi-stressor impacts on fungal diversity and ecosystem functions in streams: natural  
725 vs. anthropogenic stress. *Ecology*, **96**, 672-683.

726 Turcotte, M. M., & Levine, J. M. (2017). Phenotypic Plasticity and Species Coexistence.  
727 *Trends in Ecology & Evolution*, **31**, 803-813.

728 Van den Meersche, K., Middelburg, J. J., Soetaert, K., van Rijswijk, P., Boschker, H. T. S., &  
729 Heip, C. H. R. (2004). Carbon-nitrogen coupling and algal-bacterial interactions during  
730 an experimental bloom: Modeling a <sup>13</sup>C tracer experiment. *Limnology and*  
731 *Oceanography*, **49**, 862-878.

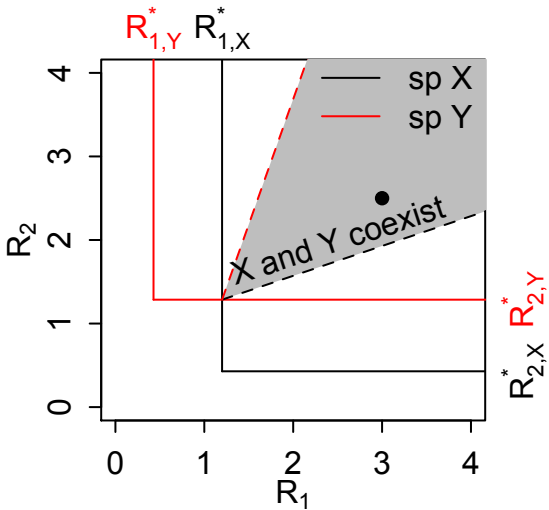
732 Vanelander, B., De Wever, A., Van Oostende, N., Kaewnuratchadasorn, P., Vanormelingen,  
733 P., Hendrickx, F., Sabbe, K., & Vyverman, W. (2009). Complementarity effects drive  
734 positive diversity effects on biomass production in experimental benthic diatom  
735 biofilms. *Journal of Ecology*, **97**, 1075-1082.

736 Vinebrooke, R. D., Cottingham, K., Scheffer, M., Norberg, J., Dodson, S., Maberly, S. C., &  
737 Sommer, U. (2004). Impacts of multiple stressors on biodiversity and ecosystem  
738 functioning: the role of species co-tolerance. *Oikos*, **104**, 451-457.

739 Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., & Garnier, E. (2007).  
740 Let the concept of trait be functional! *Oikos*, **116**, 882-892.



741 Vitousek, P. M., Aber, J. D., Howarth, R. W., Likens, G. E., Matson, P. A., Schindler, D. W.,  
742 Schlesinger, W. H., & Tilman, D. (1997). Human alteration of the global nitrogen cycle:  
743 sources and consequences. *Ecology*, **7**, 737–750.  
744



Single driver

Single driver

Both drivers

